Using Natural-Product-Based Treatments, Such as Polymer loaded with Ginger for the Management of Osteochondral Disorder

G. BEN SALAH[1,*]

[1]Department of pharmacology and toxicology, College of Pharmacy, Qassim University, Al Qassim, Saudi Arabia.

***Corresponding author**

G.SALAH@qu.edu.sa

Department of pharmacology and toxicology,

College of Pharmacy

Qassim University, Al Qassim, Saudi Arabia.

[DOI: 10.22068/ijmse.3429]

Abstract

This study reported the biological changes occurring after γ-irradiation of *in vivo* rat model and the osteochondral protective effect of Gelatine-Chitosan-Ginger (GEL-CH-GING). The results showed that Electron Paramagnetic Resonance (EPR) Spectroscopy of GEL-CH-GING showed two paramagnetic centres which correspond to g=2.19 and g= 2.002. The Fourier transform infrared spectroscopy (FTIR) analyses revealed an increase in peak intensity at C–H chains, as well as, C=O carbonyl groups. The X-ray diffraction (XRD) analysis showed no change of crystallinity. After gamma ray exposure, the rat groups have received an osteochondral defect and then were treated with GEL-CH-GING composite. Sixty days post-surgery, a significant reduction in thiobarbituric acid-reactive compounds (TBARs) was seen when compared to non-implanted rat group. Concerning oxidative stress status, GEL-CH-GIN significantly improved Superoxide Dismutase (SOD) 76 nmol/l, Catalase (CAT) 0.79 nmol/l, and Glutathione Peroxidase (GPx) 1.77 nmol/l activities in osteochondral tissue. Regarding the histomorphometric parameters of cartilaginous tissue (nCg.Th, µm), (cCg.Th, µm), (Cg.Th, µm), irradiated-GEL-CH-GIN group showed a significant increase as compared to irradiated group with 116, 74 and 188 µm, respectively (p<0.01). The microanalysis showed a high percentage of O and C in the regenertaed osteochondral tissue and indicated the deposition of novel collagen matrix**.** The biomechanical behaviour showed a significantly enhanced hardness measurement (1.73±0 .029 VH, p<0.05) when compared with that of irradiated group

Biochemical markers suggested an osteocartilage repair capacity. In fact, the levels of IL-1β, IL-6, TNF-α and VEGF in the implanted rat with GEL-CH-GING composite exhibited 51±3.48, 30.05±5.18, 65.12±4.33 and 40.42±3.32 ng/l, respectively. Our findings suggested that GEL-CH-GING composite might have promising potential applications for cartilage healing.

Key Words: Ginger, γ-irradiation, Biomaterial, oxidative stress, osteochondral defect.

[DOI: 10.22068/ijmse.3429]

1. Introduction

Irradiation may cause active deterioration and reduces matrix synthesis in bone and articular cartilage [1]. The effect of irradiation on proliferative chondrocytes illustrated the alterations in proliferation, cellular differentiation, and induction of chondrocyte apoptosis [2]. Irradiation of cartilaginous tissue enhanced proteoglycan degradation that might increase free radicals induction [3]. In fact, it has been reported that the oxidative stress derived from radiation exposure damages DNA and changes the amount and quality of cartilage cell [4]. Additionally, oxidative stress contributes to the production of inflammatory mediators like cytokines in chondrocytes [5]. The irradiation of cell raises the secretion of inflammatory cytokines, such as IL-6 as a consequence of DNA damages signalling [6].

Failure in conventional treatment of cartilage diseases has led researchers to find alternatives in tissue engineering. Recently, researchers have explored the potential of biomaterials in treating cartilage diseases. For this purpose, Chitosan (CH) is considered as a very attractive polymer and a poly-functional (poly amino-saccharide) with outstanding activities related to biodegradability, biocompatibility, antibacterial activity, low immunogenicity and controlled release behaviour [7-11]. However, the poor mechanical qualities of CH can cause hindrance to repair the damaged tissue. In fact, inadequate mechanical properties can delay the regeneration of damaged tissue and also may affect the cells viability which suggests the significance of selecting appropriate biomaterials as scaffolds for cell adhesion and proliferation.

The gelatine (GEL), a collagen-derived substance, has been shown to have attractive mechanical properties associated with the biocompatible properties with cartilaginous tissues [12,13]. GEL does not trigger any immune response in the human body [14]. It is made up of proline, hydroxyproline and glycine which help cells to join together [15]. Both CH and GEL serve as polymeric cartilage materials increasing the stability of some bioactive substance such

[DOI: 10.22068/ijmse.3429]

as polyphenol extract. In fact, this latter substance incorporation serves to protect cartilage from gamma rays due to its possessing antioxidant and anti-inflammatory properties.

Among the natural extracts rich in bioactive compounds, ginger (GING) is considered as a complex of bioactive elements such as, gingerols, shogaols, and parasols known by their anti-inflammatory activities [16]. In addition, GING extract efficiently inhibits the expression of cytokines, such as TNF-α, interleukin-6, and interleukin-8 mRNA rate. Moreover, GING significantly inhibited the production of NO and prostaglandin E-2 in cartilage tissue, which approve reducing cartilage inflammations and degradation [17].

Here, the impact of gamma irradiation on osteochondral defect was studied using *in vivo* rat model. As an alternative therapy for cartilaginous disorder, an innovative GEL-CH-GING composite was formulated and grafted as a promising therapeutic implant. To the best of our knowledge, biopolymers based GING extract has been explored for the first time to generate new osteocartilaginous tissue.

2. Materials and Methods

2.1 Preparation of GEL-CH-GING composite

The GEL solution with concentration of 5.0 wt % were prepared by dissolving 5 g of GEL powder in 100 ml distilled water for 30 min and then heated at 50 °C for 30 min under continuous stirring. CH (85% DD, degree of deacetylation) was purchased from Tunisia pharmacy. The Ginger (*Zingiber Officinale*) extract was dissolved in (2 % w/w) ethanolic solution then added gently to the solution. All mixtures were warmed and stirred at 50 °C for 30 min to obtain a homogenous solution. Finally, the mixture was prepared through repetitive freeze-thawing for four cycles.

[DOI: 10.22068/ijmse.3429]

2.2 Sterilization of GEL-CH-GING composite

The irradiations of GEL-CH-GING composite was performed at the Cobalt-60 gamma irradiation facility with energies of 1.173 and 1.332 MeV at a dose rate of 36 Gy/min. The dose rate was determined using Fricke dosimeter chemical standard dosimeter. The traceability to Aerial, the Secondary Standard Dosimetry Laboratory (SSDL), was established using the Alanine/EPR dosimetry system. The composite was placed in a polystyrene phantom to ensure electronic equilibrium and was irradiated at room temperature (293–298 K) with a dose of 15 kGy.

2.3 Electron Paramagnetic Resonance (EPR) Spectroscopy

The electron paramagnetic resonance (EPR) spectra of the composite samples were recorded at room temperature on a Bruker ER-200D spectrometer operating at 9.8 GHz X-Band frequencies with modulation amplitude of 0,2 mT , modulation frequency of 100 khz, sweep width of 210 mT and microwave power of 63 mW.

2.4 The X-ray diffraction (XRD)

The X-ray diffraction analysis of the composites were conducted using Brucker D8 advance with Cu-Kα radiation of wavelength $\lambda = 1.541$ Å in 2θ values in the range of 15–90°. The results obtained by X-ray measurement were analyzed with the X'PertHigh Score Plus program.

2.5 Fourier transform infrared (FTIR) spectroscopy

The Fourier transform infrared (FTIR) spectroscopy is used to study the structural and chemical properties of the composites. The measurement was recorded by Vertex 70 infrared spectrometer from 4000 to 400 cm^{-1} at a spectral resolution of 2 cm^{-1} and 32 scans

2.6 Rats Manipulation and Ethical Considerations

In the present study, 15–16 weeks old *Wistar* rat male (n= 40) produced in the Central Animal House were employed. The animals were given access to water *ad libitum* (Mica,

[DOI: 10.22068/ijmse.3429]

Tunisia). The animals were kept in a controlled environment with conventional temperature and humidity levels (22 2 °C and 55 5 percent, respectively), as well as, a 12-hour cycle of light and darkness. Prior to the experiment, all rats had a week of acclimatization. The rats were checked daily for clinical lameness or other complications. This study followed the guidelines for the care and use of the National Center for Sciences and Nuclear Technologies (Tunisia) which refers to Guide for the Care and Use for Laboratory Animals and was approved by the local Ethical committee of laboratory of Energy and Matter Research Laboratory (LR16CNSTN02).

2.7 Protocol of Rat Gamma-Ray Exposition

Anesthesia was induced with 10 mg/kg of ketamine (KetaminoL, Intervet International GmbH, Unterschleibheim, Germany) and 0.1 mg/kg of Xylazine (Rompun, Bayer Healthcare, Puteaux, France). Supplemented local anesthesia was applied after 25 min using 4 mg/kg carprofen (Rimadyl, Pfizer, France). Irradiation with gamma ray was established with anterior and posterior areas of photons with Cobalt 60. The dosimetric measurements for rats were performed by using a scanner. The animals were introduced in a filled packet to guarantee dose homogeneity throughout the interest area. Varian Eclipse software was used to estimate the doses of 1.5 Gy of ^{60}Co radiations in the osteochondral level in order to validate the irradiation technique. All rats were arbitrarily divided into 4 different groups with (6 animals per group) (Figure 1)

2.8 Surgical and postoperative protocols

Articular osteo-cartilaginous tissues were used as the recipients of the GEL-CH–GING composite. Right stifle joint was exposed by medial parapatellar arthrotomy after skin preparation and sterilizing, and the patella was subluxed to enable full exposure of the stifle joint. A chondral defects, 2 mm in diameter and 2 mm in height, were introduced in medial and lateral femoral condyles. The GEL-CH-GING composite were implanted into the defect area

[DOI: 10.22068/ijmse.3429]

over the medial femoral condyle. After implantation into the defects, the composite re-expanded due to the influx of fluid from bone marrow and joints and fit the chondral defect site firmly.

2.9 Tissue preparation and stress oxidative

a. Thiobarbituric acid-reactive substance (TBARS) measurements

The cartilaginous tissues were removed and rapidly frozen by dry ice. The tissues from each group were chopped, homogenized (100 mg/ml), and centrifuged at 3,000 g for 15 min at 48°C in 0.1 mol/l Tris-HCl buffer pH 7.4. By testing TBARS, which are the end result of lipid peroxidation, the amount of lipid peroxidation in the tissue homogenate was calculated [18].

b. Antioxidant enzyme studies

Superoxide dismutase (SOD) activity was measured using a spectrophotometric technique [19]. The technique used to evaluate the activity of glutathione peroxidase (GPx) is cited by Pagila and coll. [20]. The calorimetric measurement of catalase (CAT) activity at 240 nm was quantified as the number of moles of H_2O_2 used per minute per milligram of protein [21]. The method of Lowry and coll. [22] was used to measure the total protein level using bovine serum albumin.

2.10 Hardness measurement

Vickers Tester was used to gauge the hardness. The Vickers hardness (VH) number is given by the formula: H= 2P sin(h/2)/D2, where P is the applied load in kilograms, D is the average diagonal length in millimetres, and h is the angle (136°) between the Diamond Pyramid's. Measurements are taken following the composites cartilage implantation.

2.11 Histomorphometric evaluation

The implanted osteocartilaginous tissue was removed, preserved in formalin (Burdack), and then chilled. The specimens were dehydrated with alcohol solutions ranging from 70% to 100% EtOH. After that, the specimens were immersed in a solution of methyl methacrylate (MMA) and glycol methacrylate (GMA), without first being decalcified, and allowed to

[DOI: 10.22068/ijmse.3429]

polymerize. Sections of 6-7mm thick were produced using a sliding microtome (Reichert-Jung) by cutting along a transverse plane. The sections were also stained using a modified Goldner trichrome. The histomorphometric parameters were assessed using a 25-point integrating filter and the point count method [23]. From the histological zones of the implanted cartilage tissue, the subsequent parameters of cartilage thickness were determined independently and expressed as mean distances: 1-Non-calcified cartilage thickness (nCg.Th). 2-The calcified cartilage thickness (cCg.Th). 3-Total cartilage thickness (CgTh).

2.12 Measurement of Biochemical Biomarker in Serum

Venous blood draws were performed to acquire serum, which was then utilized to quantify the levels of IL-1, IL-6, TNF-, and VEGF by ELISA in accordance with the manufacturer's recommendations.

2.13 Microanalysis of implanted osteochondral tissues

The microanalysis of the implanted osteochondral tissues samples were investigated using a FEI Quanta 200 environmental scanning electron microscopy (SEM) coupled with EDAX and operating at 20 kV. The GEL-CH-GING implanted cartilage were prefixed with 2.5% glutaraldehyde solution (phosphate buffer solution, pH 7.3) overnight, and then washed with phosphate buffer solution (pH 7.4). Then, the samples were post fixed with 2% osmic acid solution (phosphate buffer solution, pH 7.4) for 90 min and dehydrated with an alcohol evaporating system. The grafted tissues were then freeze-dried with a freeze-dryer, and the process was carried out with a vapour deposition system.

2.14. Statistical analysis

All measurements are presented as means (n = 6) ± standard deviations (SD). Multiple comparisons were completed using analysis of variance (ANOVA) followed by Tukey’s range test. The probability value of $p < 0.05$ was considered significant.

[DOI: 10.22068/ijmse.3429]

3. Results

3.1 Electron Paramagnetic Resonance Analysis (EPR)

EPR detects unpaired electrons, such as the ones present in free radicals (Figure 2). When GEL-CH-GING composite was irradiated with the aim of being sterilized, free radicals are generated, from the main signal and were associated with the radicals produced by irradiation in polymer composite. Non-irradiated samples reveals two EPR signals with a *g factor* of 2.004 and 2.009 ascribed to free radicals and were associated with the radicals produced in GING cellulose containing powder [24]. Whereas in irradiated samples exhibit only one central siglet EPR signal whose intensity decreased upon irradiation with the degenerescence of the two side peaks that induced next polymer irradiation.

3.2 Fourier transform infrared (FTIR) of GEL-CH-GING composite

FTIR spectrum of control and sterilized composite with 15 kGy gamma rays dose were signalled at the absorption region of 400–4000 cm^{-1}. It is related to the tensile vibrations of the acid hydroxyl groups of GING and amine of the CH and GEL polymeric bed. In this vibration region, a band at about 3436 cm^{-1} can be the result of the hydrogen bonding formation between the GING structures and polymeric bed in the formulated composite. The bands at 784 cm^{-1} are attributed to N–H were more accentuated for the irradiated composite. The characteristic band of the transformed groups does not detect any new bands which confirm to the constriction of hydrogen bonding stretch after irradiation (Figure 3).

3.3 X-ray diffraction (XRD)

The XRD pattern showed different peaks attributed to the control and sterilized composite GE-CH-GING (Figure 4). XRD pattern of GEL powder showed amorphous morphology with a characteristic broad hump in the range of 15-30 (2θ). These characteristic

[DOI: 10.22068/ijmse.3429]

peaks are usually assigned to the triple helical crystalline structure in GEL [25]. The XRD reflection of CH revealed a sharp crystallographic peak at 22(2θ) [26]. The amorphous structure of GING cellulose also signaled a peak at around 38 (2θ) [27]. Peak intensity did not change significantly and the crystallinity of cellulose was not disrupted at large. Irradiation at 15 kGy did not transform amorphous structure via the breaking of intermolecular chemical bonds. Also, no new functional peaks of semi-crystalline composite were appeared.

3.4 Oxidative damage in the cartilage tissue

The evaluation of the oxidative stress biomarker as well as the antioxidant enzyme activity after the GEL-CH-GING implantation in the articular osteochondral tissue are shown in Figure 5. MDA level in the osteochondral tissue following 7 days of graft were significantly different when compared with that of control CT ($p< 0.05$). CAT, SOD, and GPx activities in the rat osteochondral tissue exhibited a highly significant decline when compared with those of CT rat tissues ($p< 0.05$). However, after 60 days of biomaterial implantation, a significant enhanced enzyme activities were observed in the regenerated tissue when compared with those of IR group ($p<0.05$). In fact, the SOD, CAT and GPx activities in osteochondral tissue, showed 76, 0.79, and 1.77 nmol/l, respectively.

3.5 Histological analysis

The formulated implant is adapted for implantation at an osteochondral e site to promote the growth or formation of cartilaginous tissue (Figure 6a). To assess the biomaterial effect on the osteochondal property, the explanted grafts were examined by histological analysis 60 days post-surgery. The GEL-CH-GING composite induce newly osteocartilagenous formed tissue with a marginal spreading cells (Figure 6b). The GEL-CH-GING grafts showed similar outcomes comparable to the control 60 days post-surgery (Figure 6d), the development of the

[DOI: 10.22068/ijmse.3429]

graft towards hyaline was detected without body inflammatory reaction due to the safety of the polymeric implant. This result is associated to the biocompatibility of this biomaterial and the capacity to enhance cartilage remodelling. As shown, the synthesized composite enhanced chondrogenesis that favour surface cell configuration and high cell density (Figure 6h). Histologically, a matrix-rich tissue with uniform cell distribution could be demonstrated. The staining showed the deposition of homogenous distributed proteoglycans within the matrix (Figure 6l). After 60 days, the grafts showed a positive collagen fibril assembly surrounding cells and proteoglycans.

3.6 Histomorphometric Evaluation

Histomorphometric analysis data are shown in Table 1. After 15 and 30 days of implantation, no amelioration of the histomorphometric parameters (nCg.Th, cCg.Th and Cg.Th) were detected. However After 60 days of implantation, the condylar cartilage compartments showed a significant increase of all parameters in the injured sites of cartilaginous tissues compared to that of IR rat group. In fact, the parameters nCg.Th, µm, cCg.Th, µm and Cg.Th in the irradiated-GEL-CH-GIN group showed a significant increase as compared to irradiated group with 116 , 74 and 188 µm, respectively (p<0.01). So, the GEL-CH-GING composite have the capacity to reconstruct the cartilage thickening found when comparing with the untreated-control group.

3.7 Microanalysis of implanted osteochondral tissues

The results showed the mineral distribution in the implanted GEL-CH-GING composite 60 days post-surgery (Figure 7). A high percentage of O and C were detected in the osteochondral tissue. The later indicate the novel deposition of an organic matrix made by collagen as described previously in Chen' study [28]. The presence of S may represent sulfate groups in the organic matrix. In addition, in the regenerated cartilage the other element such as

[DOI: 10.22068/ijmse.3429]

Na, K, Fe and Mg represent the release of the degraded and absorbed biomaterial in the irradiated rats.

3.8 Hardness measurement

The Table 2 illustrates the measurements hardness of the implanted osteocartilagenous tissue after 7, 15, and 60 days. The IR-GEL-CH-GING implanted tissue hardness values exhibited a similar biomechanical behavior as compared to that of native condylar cartilage tissue. In fact, this mechanical property was increased progressively and went up to even higher levels after 60 days and it reached 1.7±0.029 Vicker's hardness (VH) in the IR-GEL-CH-GING treated cartilage rat.

3.9 Measurement of Biochemical Biomarker in Serum

The serum level measurements in the different treated rats are shown in Figure 8. The levels of IL-1β, IL-6, TNF-α and VEGF in the implanted rat with GEL-CH-GING composite were 51±3.48, 30.05±5.18, 65.12±4.33 and 40.42±3.32 ng/l, respectively. These values were decreased with statistically significant differences (p<0.01) as compared with IR rats. However these biomarkers never reach those of control because the irradiation concerned all the rat body.

4. Discussion

Polymers derived from natural sources have great potential for biomedical applications. In the current study, an innovative bioactive biopolymer based on GEL, CH and GING extract was developed and characterized for osteocartilagenous graft tissue. To the best of our knowledge, no available studies explored the protective and antioxidant activity of natural polymer based GING against radiation induced harmful effects in the rat cartilage tissue.

The physico-chemical analysis of the sterilized biomaterial exhibited an elevated peak signal assigned to GEL 15-30, 22 and 38 (2θ) for both CH and GING powders, respectively. After sterilization the angle locations of GING extract interacted with CH and Gel powders were unaffected. The results specified that there was no difference in the protein backbone

[DOI: 10.22068/ijmse.3429]

structures as compared with non-sterilized composite. The peaks around 22. 2θ in the pattern proteins as noticed X-ray reflection belong to the β-sheet structure [29]. For that it was suggested that a part of crystalline cellulose derived from GING was not disrupted by bending with CH and GEL matrix [29]. On the other hand, the EPR spectra of un-irradiated GEL-CH-GING and that treated with 15 kGy was shown a doublet signal, attributed to the presence of the radiation of the radicals derived from carbohydrate. So, the analysis showed the same composition, but with different intensity ratios of the central line assigned to cellulose radicals derived from GING [30].

Concerning FTIR spectrum, the intermolecular interactions between CH and functional groups of both GEL and GING extract were established. Different functional groups such as C=C aromatics content, –OH band at wavelength of 1380 cm^{-1} were detected. The characteristic absorption bands corresponding to 3284.92 cm^{-1} are due to the -OH groups of phenolic compounds and flavonoids present in GING and the CH and GEL amine in the polymeric bed. FTIR spectrum results concluded that there was an interaction between CH, GEL matrix and GING extract during incorporation process. Moreover, the characteristic absorption bands corresponding to the vibration frequencies of 1643.64 cm^{-1} are due to the carbonyl groups of phenolic compounds [31]. The characteristic absorption bands corresponding to the vibration frequencies of 1084.86 cm^{-1} due to the OH bending of the phenolic compounds and flavonoids [32]. The bands at 784 are attributed to N–H derived from GEL matrix. However, there was no new peak significantly changed due to gamma sterilization. Containing various active functional groups, GEL-CH-GING may participate to diverse compounds which might be responsible for the various biological activities such as antioxiative phenomenon.

The rats group received ionizing radiation is subjected to the increased production of reactive oxygen species (ROS), and, as a result, an oxidative stress. Lipid peroxidation by TBARS test demonstrated that the irradiated group presents the highest level. This analysis is

[DOI: 10.22068/ijmse.3429]

an attack of various cell components by free radicals leading to biochemical changes and macromolecule modifications [33, 34]. Thus, the present study demonstrated that harmful oxidative molecules such as MDA are produced during ionizing radiation reaction at 1.5 Gy of the whole body irradiation of rats.

ROS have a harmful effect on the cellular antioxidant defence mechanisms by falling the level of antioxidant enzymes in osteocartilaginous tissue, especially catalase (CAT), superoxide dismutase (SOD) and glutathione peroxidase (GPx). The formulated biomaterials contain natural antioxidants derived from GING that were applied to detoxify the increased levels of free radicals. In fact, after the GEL-CH-GING graft, the antioxidant mechanisms of the body were enhanced as shown by elevated cartilage GP_X, SOD and CAT activities with reduction of cartilaginous MDA level.

These results might be attributed to the reaction between the free radicals and the free residual amino groups to form CH ammonium groups. In addition, it might be attributed to the anti-oxidative effect of phenolic compounds, such as quercetin, zingerone, gingerenone-A, and 6-dehydrogingerdione [35]. Morever, as described, GING extract contains several terpene components, such as β-bisabolene, α-curcumene, zingiberene, α-farnesene, and β-sesquiphellandrene known as very powerful antioxidant components [35]. The role of antioxidants is to lower or terminate these chain reactions by removing free radicals or inhibiting other oxidation reactions [36].

On the other hand, our data revealed elevated serum level of IL-1β, IL-6, TNF-α and VEGF indicating that the irradiation induced inflammatory processes. This finding has been previously reported in Lee's study that demonstrated that irradiation marked up-regulation of mRNA and protein expression of the pro-inflammatory mediator TNF-α [36]. After GEL-CH-GING graft, these biomarkers were reduced but did not return to the normal level. This finding might be explained by the fact that the reparation is focusing in the focal areas of degenerated

[DOI: 10.22068/ijmse.3429]

cartilage however the irradiation concerned all the body. Moreover, it has been reported that the bioactive ingredients in GING, such as gingerols, shogaols exert anti-inflammatory protective effects on chondrocytes and human synoviocytes by specifically decreasing the production of inflammatory mediators [37]. In addition, gingerol suppresses inflammatory degradation enzymes such as the nitric oxide (NO) synthase, which has been shown to be regulated and promoted by the master transcription factor NF-κB [37]. Also, GING suppress the expression of pro-inflammatory cytokines, such as TNF-α, interleukin-6, and interleukin-8 mRNA levels, thereby reducing cartilage inflammations and degradation [37].

The microanalysis of the implanted cartilage revealed that the organic matrix after remodelage and some of acid groups might possibly derived from sulfate groups of glycosaminoglycans obtained from CH. In fact, in cartilage matrix, virtually all the sulfur is found as sulfate in (Glycosaminoglycans) GAG [37,38]. Also, it is well known that CH is composed of glucosamine and N-acetyglucosamine with structure and characteristics similar to GAG [38]. Thus, the CH exhibits multiple bioactivities, such adapted microenvironment for native cells to adhere, colonize, and repair the damaged; however, it does not promote chondrocytes differentiation, proliferation, and matrix secretion [39].

The GEL-CH-GING biomaterial can provide the properties not found in CH matrix [40]. In fact, due to its similarity to collagen, GEL can promote cell adhesion, differentiation, and proliferation. In addition GEL was providing enough surface area for chondrocyte attachment, matrix production, and accumulation and biocompatibility. Furthermore, GEL-CH-GING was biodegradable, and the rate of degradation matches the rate of chondrogenesis.

The biomechanical behaviour of GEL-CH-GING composite enhanced the insufficient mechanical qualities of cartilage caused by irradiation mainly on proteoglycan production. In fact, it is one of the primary components of cartilage matrices production, playing a key role in providing osmotic resistance to resist compressive load [41,42]. After 60 days, no significant

[DOI: 10.22068/ijmse.3429]

difference was found between the experimental specimens and the native cartilage. In the beginning of cartilage remodelage, clusters of neocartilage were observed. Then, the biomaterial gradually underwent degradation, and the tissues developed further, matured, and formed relatively homogeneous cartilage, by 60 days a mature chondrocytes were found embedded in lacuna within the tissue-engineered cartilage.

The inhibition of oxidative damage by incorporation of strong antioxidants such as GING extract into GEL and CH matrix becomes an attractive therapeutic strategy, and the present formulated biomaterial may predict an expected clinical effect for osteocartilagenous tissue disorder following radiation-induced complications.

5. Conclusion

Overall, this study demonstrated that GEL-CH-GING composite could protect osteochondral tissue, enhance the biomechanical property, considerably ameliorate oxidative stress balance and alleviate the destructive effects of ionizing radiation. Thus, the synthetised composite is useful as a radioprotective for osteocartilagenous tissue. These beneficial effects of ginger may be due to its antioxidant properties. It may be considered as a natural product to prevent osteochondral cartilage destruction in the clinical setting.

Acknowledgments

The researcher(s) would like to thank the Deanship of Scientific Research, Qassim University for funding the publication of this project.

References

1. Jebahi, S., Saoudi, M., Farhat, L., Oudadesse, H., Rebai, T., Kabir, A., El Feki, A., & Keskes, H., *"Effect of novel curcumin-encapsulated chitosan-bioglass drug on bone and skin repair after gamma radiation: experimental study on a Wistar rat model". Cell biochemistry and function, 2015, 33(3), 150–159.* https://doi.org/10.1002/cbf.3098

[DOI: 10.22068/ijmse.3429]

2. Saintigny, Y., Cruet-Hennequart, S., Hamdi, D.H., Chevalier, F., & Lefaix, JL., *"Impact of therapeutic irradiation on healthy articular cartilage". Radiation research, 2015, 183(2), 135–146.* https://doi.org/10.1667/RR13928.1
3. Zahan, O.M., Serban, O., Gherman, C., & Fodor, D. *"The evaluation of oxidative stress in osteoarthritis". Medicine and pharmacy reports, 2020, 93(1), 12–22.* https://doi.org/10.15386/mpr-1422
4. Dai, S., Wen, Y., Luo, P., Ma, L., Liu, Y., Ai, J., & Shi, C., *"Therapeutic implications of exosomes in the treatment of radiation injury". Burns & trauma, 2022, 10, tkab043.* https://doi.org/10.1093/burnst/tkab043
5. Choi, S.Y., Kim, S., & Park, K.M., *"Initial Healing Effects of Platelet-Rich Plasma (PRP) Gel and Platelet-Rich Fibrin (PRF) in the Deep Corneal Wound in Rabbits". Bioengineering (Basel, Switzerland), 2022, 9(8), 405.* https://doi.org/10.3390/bioengineering9080405
6. Cinat, D., Coppes, R.P., & Barazzuol, L., *"DNA Damage-Induced Inflammatory Microenvironment and Adult Stem Cell Response". Frontiers in cell and developmental biology, 2021, 9, 729136.* https://doi.org/10.3389/fcell.2021.729136
7. Jebahi, S., Odadesse, H., Saoudi, M., Kallabi, F., Pellen, P., Rebai, T., El Feki, A., Keskes, H., *"Cytocompatibility, gene-expression profiling, apoptotic, mechanical and (29) Si., (31)P solid-state nuclear magnetic resonance studies following treatment with a bioglass-chitosan composite*". *Biotechnology letters, 2014, 36(12), 2571–2579.* https://doi.org/10.1007/s10529-014-1633-z
8. Jebahi, S., Ben Salah, G., Saoudi, M., Besaleh, S., Oudadesse, H., Mhadbi, M., Rebai, T., Keskes., H., El Feki, A., *"Genotoxicity effect, antioxidant and biomechanical correlation: experimental study of agarose-chitosan bone graft substitute in New Zealand white rabbit model". Proceedings of the Institution of Mechanical Engineers.*

[DOI: 10.22068/ijmse.3429]

Part H., Journal of engineering in medicine, 2014, *228(8), 800–809.* https://doi.org/10.1177/0954411914547247

9. Jebahi, S., Oudadesse, H., Ben Salah, G., Saoudi, M., Mesadhi, S., Rebai, T., Keskes, H., El Feki, A., and El Feki, H., *"Chitosan-Based Bioglass Composite for Bone Tissue Healing: Oxidative Stress Status and Antiosteoporotic Performance in Ovariectomized Rat Model". Korean Journal of Chemical Engeneering, 2014, 31, 1616-1623.* https://doi.org/10.1007/s11814-014-0072-9
10. Jebahi, S., Bessalah, S., Zagrouba, M., Hajji, S., Raoufi, A., Hidouri, M., *"The Effect of Gamma Rays on Gelatin / Chitosan Wound Dressing Physico-Chemical and Biological Properties". Iranian Journal of Materials Science and Engineering, 2022, 19, 2.*
11. Jebahi, S., Bessalah, S., Raouafi, A., Hajji, S., Keskes, H., Hidouri, M., *"Novel bioactive adhesive dressing based on gelatin/ chitosan cross-linked cactus mucilage for wound healing". The International journal of artificial organs, 2022, 45(10), 857–864.* https://doi.org/10.1177/03913988221114158
12. Martinez-Garcia, F.D., Van Dongen, J.A., Burgess, J.K., Harmsen, M.C., *"Matrix Metalloproteases from Adipose Tissue-Derived Stromal Cells Are Spatiotemporally Regulated by Hydrogel Mechanics in a 3D Microenvironment". Bioengineering (Basel., Switzerland), 2022, 9(8), 340.* https://doi.org/10.3390/bioengineering9080340
13. Sulaiman, S.B., Idrus, R., Hwei, N.M., *"Gelatin Microsphere for Cartilage Tissue Engineering: Current and Future Strategies". Polymers, 2020, 12(10), 2404.* https://doi.org/10.3390/polym12102404
14. Bessalah, S., Jebahi, S., Raoufi, A., Faraz, A., Zagrouba, M., Hammadi, M., *"Physicochemical Evaluation and in Vitro Hemocompatibility of Goat Gelatin: Bioactive Dressing to Promote Wound Healing". Iranian Journal of Materials Science and Engineering, 2022, 19(2).* https://doi.org/10.22068/ijmse.2599

[DOI: 10.22068/ijmse.3429]

15. Li, P., Wu, G., *"Roles of dietary glycine, proline, and hydroxyproline in collagen synthesis and animal growth". Amino acids, 2018, 50(1), 29–38.* https://doi.org/10.1007/s00726-017-2490-6

16. Mao, Q.Q., Xu, X.Y., Cao, S.Y., Gan, R.Y., Corke, H., Beta, T., Li, H,B., *"Bioactive Compounds and Bioactivities of Ginger (Zingiber officinale Roscoe)". Foods (Basel, Switzerland), 2019, 8(6), 185*. https://doi.org/10.3390/foods8060185

17. Leyva-López, N., Gutierrez-Grijalva, E.P., Ambriz-Perez, D.L., Heredia, J.B., *"Flavonoids as Cytokine Modulators: A Possible Therapy for Inflammation-Related Diseases". International journal of molecular sciences, 2016, 17(6), 921.* https://doi.org/10.3390/ijms17060921

18. Buege, J.A., Aust, S.D., *"Microsomal lipid peroxidation". Methods Enzymol., 1984, 105, 302–310.*

19. Marklund, S., Marklund, G., *"Involvement of the superoxide anion radical in the autoxidation of pyrogallol and convenient assay for superoxide dismutase". Eur J Biochem., 1975, 47, 469–474.*

20. Pagila, D.E., *"Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase". J Lab Clin Med., 1967, 70, 158–169.*

21. Aebi, H., *"Catalase in vitro". Methods Enzymol. 1984, **105**, 121–126.*

22. Lowry, O.H., Rosebrough, N.J., Farr, AL., et al. "Protein measurement with Folin phenol reagent". *J Biol Chem.* 1951, 193, 265–275.

23. Schenk, R.K., Merz, W.A., Müller, J.A., *"Quantitative histological study on bone resorption in human cancellous bone". Acta Anat. 1969, 74, 44-53.*

24. Ahn, J.J., Akram, K.J.D., Kwon, J.H., *"Investigation of Different Factors Affecting the Electron Spin Resomance-based Characterization of Gamma-irradiated Fresh, White,*

[DOI: 10.22068/ijmse.3429]

and Red Ginseng". Journal of ginseng research, 2012, 36(3), 308–313. https://doi.org/10.5142/jgr.2012.36.3.308

25. Chetouani, A., Elkolli, M.M., Bounekhel, M., Benachour, D., *"physicochemical characterization of gelatin- cmc composite edibles films from polyion-complex hydrogels". J. Chil. Chem. Soc. 2014, 59(1), 2279-2283.*

26. Kumar, S., Koh, J., *"Physiochemical, Optical and Biological Activity of Chitosan-Chromone Derivative for Biomedical Applications". International Journal of Molecular Sciences, 2012, 13(5), 6102–6116.*

27. Lei, H., Wei, Q., Wang, Q., Su, A., Xue, M., Liu, Q., Hu, Q., *"Characterization of ginger essential oil/palygorskite composite (GEO-PGS) and its anti-bacteria activity". Materials science & engineering. C. Materials for biological applications, 2017, 73, 381–387.* https://doi.org/10.1016/j.msec.2016.12.093

28. Chen, J., Li, J., Li, Z., Yi, R., Shi, S., Wu, K., Li, Y., Wu, S., *"Physicochemical and functional properties of type I collagens in red stingray (Dasyatis akajei)". skin. Mar. Drugs, 2019, 17, 558.*

29. Abral, H., Ariksa, J., Mahardika, M., Handayani, D., Aminah, I., Sandrawati, N., Sugiarti, E., Muslimin, AN., Rosanti, S.D., *"Effect of heat treatment on thermal resistance, transparency and antimicrobial activity of sonicated ginger cellulose film". Carbohydrate polymers, 2020, 240, 116287.* https://doi.org/10.1016/j.carbpol.2020.116287

30. Yan, H., Li, P.H., Zhou, G.S., Wang, Y.J., Bao, B.H., Wu, Q.N., Huang, S.L., *Rapid and practical qualitative and quantitative evaluation of non-fumigated ginger and sulfur-fumigated ginger via Fourier-transform infrared spectroscopy and chemometric methods*". *Food chemistry, 2021, 341(1), 128241.* https://doi.org/10.1016/j.foodchem.2020.128241

[DOI: 10.22068/ijmse.3429]

31. Jan, R., Gani, A., Masarat, D.M., Bhat, N.A., *"Bioactive characterization of ultrasonicated ginger (Zingiber officinale) and licorice (Glycyrrhiza Glabra) freeze dried extracts". Ultrasonics sonochemistry, 2022, 88, 106048.* https://doi.org/10.1016/j.ultsonch.2022.106048

32. Saoudi, M., Jebahi, S., Jamoussi K., Ben Salah, G., Kallel, C., El Feki, A., *"Haematological and biochemical toxicity induced by methanol in rats: ameliorative effects of Opuntia vulgaris fruit extract". Human & experimental toxicology, 2011, 30(12), 1963–1971*. https://doi.org/10.1177/0960327111403175

33. Saïdi, S.A., Abdelkafi, S., Jbahi, S., van Pelt, J., El-Feki, A., *"Temporal changes in hepatic antioxidant enzyme activities after ischemia and reperfusion in a rat liver ischemia model: effect of dietary fish oil". Human & experimental toxicology, 2015, 34(3), 249–259.* https://doi.org/10.1177/0960327114531991

34. Sharifi-Rad, M., Varoni, E.M., Salehi, B., Sharifi-Rad, J., Matthews, K.R., Ayatollahi, S.A., Kobarfard, F., Ibrahim, S.A., Mnayer, D., Zakaria, Z.A., Sharifi-Rad, M., Yousaf, Z., Iriti, M., Basile, A., Rigano, D., *"Plants of the Genus Zingiber as a Source of Bioactive Phytochemicals: From Tradition to Pharmacy". Molecules (Basel, Switzerland), 2017, 22(12), 2145*. https://doi.org/10.3390/molecules22122145

35. Jebahi, S., Oudadesse, H., Feki, H., Rebai, T., Keskes, H., Pascal, P., El Feki, A., *"Antioxidative/oxidative effects of strontium-doped bioactive glass as bone graft in vivo assays in castrated rats". Journal of applied biomedicine, 2012, 10, 195–209.*

36. Lee, W.H., Sonntag, W.E., Mitschelen, M., Yan, H., Lee, Y.W., *"Irradiation induces regionally specific alterations in pro-inflammatory environments in rat brain". International journal of radiation biology, 2010, 86(2), 132–144.* https://doi.org/10.3109/09553000903419346

[DOI: 10.22068/ijmse.3429]

37. Wojdasiewicz, P., Poniatowski, ŁA., Szukiewicz, D., *"The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis". Mediators of inflammation, 2014, 561459*. https://doi.org/10.1155/2014/561459

38. Wei, Z., Zhang, G., Cao, Q., Zhao, T., Bian, Y., Zhu, W., Weng, X., *"Recent Developments and Current Applications of Organic Nanomaterials in Cartilage Repair". Bioengineering (Basel, Switzerland), 2022, 9(8), 390.* https://doi.org/10.3390/bioengineering9080390

39. Jebahi, S., Oudadesse, H., Bui, X.V., Keskes, H., Rebai, T., El Feki, A., El Feki, H., *"Repair of bone defect using bioglass-chitosan as a pharmaceutical drug: An experimental study in an ovariectomised rat model". African Journal of Pharmacy and Pharmacology, 2012, 6(16), 1276-1287*.

40. Han, E.H., Chen, S.S., Klisch, S.M., Sah, R.L., *"Contribution of proteoglycan osmotic swelling pressure to the compressive properties of articular cartilage". Biophysical journal, 2011, 101(4), 916–924.* https://doi.org/10.1016/j.bpj.2011.07.006

41. Wu, C.L., Harasymowicz, N.S., Klimak, MA., Collins, K.H., Guilak, F., *"The role of macrophages in osteoarthritis and cartilage repair". Osteoarthritis and cartilage, 2020, 28(5), 544–554.* https://doi.org/10.1016/j.joca.2019.12.007

42. Van Boxtel, J., Vonk, LA., Stevens, H.P., van Dongen, J.A., *"Mechanically Derived Tissue Stromal Vascular Fraction Acts Anti-inflammatory on TNF Alpha-Stimulated Chondrocytes In Vitro". Bioengineering (Basel, Switzerland), 2022, 9(8), 345.* https://doi.org/10.3390/bioengineering9080344

[DOI: 10.22068/ijmse.3429]